Natal homing in juvenile loggerhead turtles (Caretta caretta)

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Abstract

Juvenile loggerhead turtles (Caretta caretta) from West Affantic nesting beaches occupy oceanic (pelagic) habitats in the eastern Atlantic and Mediterranean, whereas larger juvenile turtles occupy shallow (neritic) habitats along the continental coastline of North America. Hence the switch from occanic to nertific stage can involve a trans-oceanic migration. Several researchers have suggested that at the end of the oceanic phase, juveniles are homing to feeding habitats in the vicinity of their natal rookery. To test the hypothesis of juvenile homing behaviour, we surveyed 10 juvenile feeding zones across the eastern USA with mitochondrial DNA control region sequences (N = 1437) and compared these samples to potential source (nesting) populations in the Atlantic Ocean and Mediterranean Sea (N = 465). The results indicated a shallow, but significant, population structure of neritic juveniles $(\Phi_{SY} = 0.0088, P = 0.016)$, and haplotype frequency differences were significantly correlated between coastal feeding populations and adjacent nesting populations (Mantel test R2 = 0.52, P = 0.001). Mixed stock analyses (using a Bayesian algorithm) indicated that juveniles occurred at clevated frequency in the vicinity of their natal rookery. Hence, all lines of evidence supported the hypothesis of juvenile homing in loggerhead turtles. While not as precise as the homing of breeding adults, this behaviour nonetheless places juvenile turtles in the vicinity of their natal nesting colonies. Some of the coastal hazards that affect declining nesting populations may also affect the next generation of turtles feeding in nearby habitats.

Keyatords: Bayesian, conservation genetics, marine turtles, mitochandrial DNA, mixed stock analysis

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Introduction

Loggerhead turtles have two distinct juvenile stages, the first being an oceanic stage after hatching (Carr 1987; Bulten

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2003a). For posthatchling turtles departing the nesting beaches of the western Allanie, this oreanic habitariculules waters around the Azones and Madeira, and the Grand Busis (Newformalland, Carada), as well as the Mediterranean Sen (Bolten et al. 1998; Laurent et al. 1998; hibra. 2003a; LaCassella et al. 2004; Subsequent to the occanic stage, which may span a decade (Bjorndal et al. 2009). 2003, most older juvenules enter a nertitic (benthic feeding) stage, in which they consume hard-shelled invertershear in shallow habitats of the verstern Atlantic (Dodd 1988; Bolten 2003b). Whereas the juurney from nesting benches to occaric juvenile habitat is largely mediant by passive fransport, the return trip may include active orientation and swimming (Bolten 2003a).

Upon reaching sexual maturity, female loggerheads make reproductive migrations to breed and nest in the vicinity of their natal beach (Bowen et al. 1993). Male loggerheads may make a similar migration to breeding areas near their natal beach (see FitzSimmons et al. 1997a,b). Hence homing behaviour is widely accepted for the reproductive migrations of adults. This raises the question of whother juveniles also exhibit homing behaviour during their trans-oceanic migration. Genetic markers hold considerable promise for addressing this issue. As a consequence of natal homing by females, must nesting populations are distinguished by differences in the frequency of mitochondrial DNA (mtDNA) haplotypes (Bowen et al. 1994; Encalada et al. 1998; Hatase et al. 2002a). It is possible to use these natural genetic 'tags' to resolve the origin of feeding populations, even when the feeding population is a mixture of turtles from several source nesting populations (Bowen 1995, 2003). Mixed stock analyses have been used to monitor salmon for over 20 years (Grant et al. 1980), but this methodology has seldom been applied to other organisms (Broderick et al. 1994; Epifanio et al. 1995; Wirgin et al. 1997).

Previous mtDNA studies by Norrgard & Graves (1996), Rankin-Baransky et al. (2001), Witzell et al. (2002), Engstrom et al. (2002) and Bass et al. (2004) concluded that contributions to juvenile loggerhead habitats are influenced by the size of regional source (nesting) populations. The very large rookery in southern Florida contributes most of the neriticstage turtles feeding along this coast, with smaller but significant contributions from the rookeries on the Yucatan peninsula, Gulf of Mexico, and the Atlantic states of Georgia, South Carolina and North Carolina. The same mtDNA studies prompted authors to suggest that juvenile turtles may tend to feed in the vicinity of their natal nesting colony. Hence, two factors have been postulated to explain the composition of juvenile loggerhead feeding populations: size of source populations and proximity to these sources. A third factor, male-biased dispersal, has been proposed for loggerhead juveniles in the Mediterranean (Casale et al. 2002).

These mIDNA surveys invoke the possibility that loggerhead turtles have two distinct homing migrations the well-known reproductive migration, that brings salulis back to their ancestral breeding areas, plus a juvenile migration that brings oneasite ungaratis to nertic reading habitats near their location of origin. If juvenile homing occurs in the northwestern Atlantic, how precise is this behaviour? Do older juvenile turtles return to a broad region of the western Atlantic, or do they send to recruit to beding grounds near their nockey of origin? The latter possibility



Fig. 1 Map. mdicating the location of sampled rookedes and foraging grounds in North America. For alobeginitions see Table, 1 and 2. Divisions between EL-NA and FL-SA and between FL-NG and FL-SG are indicated by dark hars at Cape Canaveral and Tampa Bay, respectively.

thoming towards natal location) would carry the expectation of population genetic differences among regional feeding cohorts. In parallel with the genetic differences observed between nesting populations (Bowen et al. 1993). Psevious studies invoked the possibility of juverile horing (Norgard & Graves 1996; Rankin-Baransky et al. 2001; Engstrom et al. 2012; Witsell et al. 2002). However, all of these survegs examined a single feeding habitat. To evaluate the venezity and precision of homing, samples from multiple feeding leations across the vestor. Atlantic are necessary.

Here we survey the mIDNA control region sequences of 1457. neritie-freeting individuals (strandings and live captures) along the Atlantic coost of North America from Texas to New England (Fig. 1). We incurporate three previous surveys of juveniles from this region (Rankinsbaranskey et al. 2001; Witzell et al. 2002; Bans et al. 2004) along with complementary surveys of regional mesting lemales (Encaladard al. 1996; Fearres (2011) to resolve the origin of neritic-freeding loggerhead populations and to assess the site flieflity of juveniles. This survey is intended to resolve a gap in loggerhead life history, but the information is relevant to conservation strategies, as thousands of juvenile loggerhead furtles are caught in fishing gear and dredging operations. Wildlife managers need to know which beeeding populations are affected by these activities.

Materials and methods

Nest samples had been proviously collected from 465 females or progeny from Quintana Roo (Yucatan Peninsula, Mexico).

Table 2. Juvenile feeding populations, from biopsies in North Carolina, power plant entrapment system (n = 106) and strandings on the coast of southern Florida, and strandings from everywhere else, as described in Rankin-Baransky et al. (2001), and Witzell et al. (2002)

Haplotype	TX	PLNG	FL-5G	FL-SA	FL-NA	GA	5C	NC	- VA	NE-US
A CC-Al	67	10.	14	59	37	107	49	166	143	90
& CC-AZ	53	6	21	58	28	68	32	98	91	53
C CC-A3	10	1	5	10	1	10	5	A	4	9
o CC-A4								1		
e CC-AS						2	1	3	t	1
F CC-A6										
D OC-A7	4			2		3	. 2	5	4	1
H CC A8		4.0		1			7	7		
₹ CC-A9				4	1			9		
J CC-A10	2		3941.6%	2		1	2	4	3	A 1
K CC-ATT						1.0				
CC-A13	1					2			2	
CC-A14	5			6		10		7 3 4 T	4.0	3
CC-A18								1		4.73
CC-A19										
CC-A20			2	7						
CC-A22						738 POS				
CC-A23						i.				
Total	142	18	45	146	69	209	95	296	256	161

Full sequence descriptions are available from http://accstructl.edu/combina.html. Abbreviations: TX = Treas, H.-NG = Exercia Penius no, northern Gulf af Mexice, Ft.-SG = Florida Penius no, northern Gulf af Mexice, Ft.-SG = Florida Penius no, northern Atlantic coast, GA = Georgie, SC = South Caroline, NC = North Caroline, vC = Virginia; NE-US = northern Atlantic coast, GA = Georgie, SC = South Caroline, NC = North Caroline, vC = Virginia; NE-US = northern Atlantic coast, GA = Georgie, SC = South Caroline, NC = North Caroline, vC = Virginia; NE-US = northern Atlantic coast, GA = Georgie, Ncw York and Massachusetts.

Fisheries Science Center in La Jolla, CA. (Applied Biosystems Inc. models 377 and 3100). Chromatograms were aligned using SEQUENCHER 3.1 (Gene Codes, Inc.). These sequences were compared to previously identified haplotypes from nesting and foraging locations (Bolten et al. 1998; Eucalada et al. 1998) and were assigned haplotype numbers based on the web site maintained by the Archie Carr Center for Sea Turtle Research (http:// scstf.ulf.edu/comidina.html).

The mtDNA diversity among populations was measured with an analysis of molecular variance (AMOVA) as implemented in ARLEOUIN version 2.0 (Schneider et al. 2000). The same software package was used to conduct a Mantel test and to estimate haplotype diversity, nucleofide diversity and haplotype frequencies (Nei 1987; Excoffier & Slatkin 1995). In all tests that required estimates of sequence divergence, the Tamura-Nei model of nucleotide substitutions was employed (Tamura & Nei 1993). The Mantel test is a comparison of genetic differentiation (Dcr values) among seven nesting colonies (FL-NG, FL-SG, FL-SA, FL-NA, GA, SC, NC in Table 1) along the continental coast of North America (X-matrix), and seven proximal feeding zones (Ymatrix). The correlation between these two matrices was evaluated with a permutation test as described by Smouse et al. (1986). In a related test, the frequency of the most common haplotype (CC-A1) was compared between seven resting areas and adjacent feeding populations, rabulating the standard Prosson coefficient of determination (\mathcal{R}^3) and using a permutation test (with 30 000 permutations) to calculate two-tailed significance against a null hypothesis of \mathcal{R}^2 =0.

The availability of data from multiple mixed stocks (feeding populations) and multiple sources (nesting areas), as well as the need to account for the effects of population size, has led us to develop a variation of standard Bayesian methods for mixed stock analysis (see Pella & Masuda 2001; Bolker et al. 2003; Bass et al. 2004; Okuvama & Bolker 2005). Mixed stock analysis normally estimates the proportion of individuals in a single mixed stock contributed by each of a number of source populations. In the current analysis, where we had multiple sources and multiple mixed stocks, we computed the same parameters-the proportion of each mixed stock contributed by each mokery-but we were also interested in partitioning the other way, finding the fractions of the total contribution to the northwest Atlantic (from each source population) that were present in each mixed stock. In particular, we wanted to know if rookeries contributed disproportionately to nearby feeding populations. Estimating contributions to foraging grounds separately also assumed that all foraging grounds were the same size. Even if we could say that 57% of the feeding individuals in the northeast USA and 69% of those

in South Florida originated from the South Florida rookery, we could not necessarily infer that more turtles from South Florida go to the South Florida foreging ground, if the northern foreging ground had a larger population. Our new method did not make this assumption.

We can estimate these parameters in a straightforward way, if we know the relative sizes of the rookeries and if we make the assumption that the total contributions of nesting populations to the combined feeding populations are proportional to their size. Essentially, if the relative sizes (and therefore overall proportional contributions) of the rookeries are known, we can derive an equation to translate between the partitioning of mixed-stock turtles according to their nesting population of origin and the partitioning of nesting population turtles according to their mixed-stock destination (Bolker et al. unpublished results). This procedure can then be incorporated into standard mixed-stock estimation procedures, either in a maximum-likelihood or in a Bayesian estimation framework; we have chosen a Bayesian framework because of its more accurate calculation of confidence limits (Bolker et al. 2003). We can examine the estimates from this procedure either in the traditional way. as the fractions of each mixed stock estimated to come from each source population (rookery), or, in the new way, as the fractions of the total contribution from each rookery estimated to be present in each mixed stock. The new method also adds an 'unknown' category to the list of mixed stocks, to allow for the possibility that some of the sampled rookeries contribute to one or more unknown mixed stocks.

Designation of source populations for mixed stock analyses

Based on the population subdivisions defined by Encalada et al. (1998), Laurent et al. (1998) and Pearce (2001), the nesting samples (Table 1) were grouped into the following source populations for mixed stock analysis: (i) Florida coast in the northern Gulf of Mexico (FL-NG: n = 49); (ii) South Florida (SFL, n = 109) combining southern Atlantic and southern Gulf coasts of Florida (FL-SA and FL-SG in Table 1); (iii) northeast Florida to North Carolina (NEFL-NC; n = 105), combining northern Atlantic coast of Florida, Georgia, South Carolina and North Carolina (FL-NA, GA, SC, and NC in Table 1); (iv) Dry Tortugas, FL (DT, n = 58); (v) Quintana Roo, Yucatan, Mexico (MX, n = 20); (vi) Bahia, Brazil (BR, n = 11); (vii) Greece (GR, n = 81); and (viii) Turkey (TU; n = 11); 32). These groupings are based on statistically significant differences in haplotype frequencies. Additional population subdivisions almost certainly exist but cannot be detected with the available mtDNA sequence information.

Designation of feeding populations for mixed stock analysis fuvenile populations from Texas to the northeast USA (Table 2) were analysed without modification for indices of genetic diversity, haplotype frequency comparisons, F statistics, correlation coefficients and the Mantel test. Because of the relatively sparse data from the mixed stocks, analysing the contributions to the mixed stocks in their fully disaggregated form led to very wide confidence limits. We estimated the contributions to each of 11 'foraging grounds' (the 10 foraging grounds represented by the different data sets in Table 2 plus an unknown foraging ground) separately, but we also placed the results into four groups representing a northern mixed stock (N: FL-NA, GA, SC, NC, VA, NEUS in Table 2), a southern mixed stock (S: FL-SA in Table 2), a Gulf of Mexico mixed stock (G: TX, FL-NG, FL-SG in Table 2) and an unknown mixed stock (not shown). As well as considering the basic estimate of contribution to each of the known mixed stocks, we also considered the ratio of the contributions to the total contributions to known stocks le.g. N/(N+S+G) would represent the contribution to the Northern stock relative to the combined total of Northern, Southern and Gulf contributions. In addition, we discard 'orphan' haplotypes, the haplotypes from the feeding grounds that were not detected in nesting populations (haplotypes CC-A18, -19, -22 and -23 in Table 2): these specimens provided no additional information about the contributions of nesting populations to feeding grounds. The individuals with 'orphan' haplotypes (n = 4)comprised less than 1% of the overall feeding ground sample.

Results

In the analysis of feeding grounds and nexting populations, we encountered 18 of the 28 haplotypes reported for Atlantic loggerhead turtles (Tables 1 and 2). Haplotype diversity in juvenile feeding propulations was fairly uniform θ = 0.555–0.684) as was nucleotized diversity θ = 0.0221–0.02494). In both cases the lowest diversity estimates were from locations in Florida (Table 3).

The test of population structure and natal homing in juvenile turtles consisted of three classes of data analysis. The first was a comparison of haplotype distributions among 10 feeding zones along the Atlantic coast of North America, from Texas to Massachusetts. These feeding zones correspond to US states and federal management regimes, including Texas, four zones around the Florida peninsula, Georgia, South Carolina, North Carolina, the seasonal feeding habitat in Virginia, and the seasonal feeding habitat from Maryland to Massachusetts (Fig. 1). Results of AMOVA indicated that juvenile turtles were not randomly distributed among these regions: $\Phi_{ST} = 0.0088$. This value was low on the scale of population genetic separations, but was significant in permutation tests (P = 0.016). When just the seven feeding grounds that are adjacent to continental nesting colonies (FL-NC, FL-SG, FL-SA, FL-NA,

Table 3 Feeding habital diversity estimates, including haplotype diversity (i) and nucleotide diversity (ii)

	Haplotype	Nucleotide diversity (n)		
Habitet	diversity (h)			
TX	0.635 ± 0.025	0.02442 ± 8.01247		
FL-NG	0.608 ± 0.086	0.02475 ± 0.01330		
FL-SG	0.684 ± 0.048	0.02214 ± 0.01159		
FL-SA	0.676 ± 0.024	0.02492 ± 0.01273		
FL-NA	0.555 ± 0.032	0.02380 ± 0.01227		
GΛ	0.630 ± 0.024	0.02451 ± 0.01251		
5C	0.622 ± 0.035	0.02414 ± 0.01238		
NC	0.576 ± 0.021	0.02337 ± 0.01193		
VA	0.563 ± 0.020	0.02339 ± 0.01194		
NE US	0.579 ± 0.028	0.02355 ± 0.01205		

Abbreviations are listed in Tables 1 and 2.

GA, SC, NC in Table 2) were compared, the corresponding values were somewhat higher $\Phi_{gr} = 0.0164$ (P < 0.006). The same comparison with conventional P-statistics (which do not hickude the divergence between haplotypes) yielded a lower but significant value $(P_{gr} = 0.0070$; P = 0.035. Hence our first conclusion is $(P_{gr} = 0.0070$; P = 0.035. Hence our first conclusion is that juvenite loggerhead turtles are not distributed randomly among feeding behavior

Our second approach was an assessment of haplotype frequencies at nesting populations compared to adjacent (juvenile) feeding populations. The frequency of the most common haplotype (CC-A1) at the seven nesting populations was significantly correlated with the frequencies at the seven adjacent feeding populations ($R^2 = 0.88$, P = 0.049; Fig. 2). Given that our data matrices included only seven values, the significant outcome was especially compelling. A Mantel test of genetic distances (Φ_{ST} values) among the seven nesting colonies (X-matrix) vs. genetic distances among seven feeding populations (Y-matrix) produced highly significant results ($R^2 = 0.52$, P = 0.001). Approximately half of the genetic divergence among juvenile feeding populations is correlated to genetic divergence among corresponding nesting populations. These two correlation tests provided strong support for our second conclusion, that haplotype distributions in juvenile feeding populations. are significantly influenced by the composition of nearby nesting populations.

Third, the results of the Bayesian mixed stock malysis showed that nesting populations did indeed contribute more to neighbouring mixed stocks than to distant mixed stocks (Tables 4 and 5). For most rockeries, the data were too space to determine the destinations of neutric juveniles with certainty. The mean and median estimates of the fraction going to any one of the three flumped mixed shocks were close to proportional to the number of foreging grounds included in the mixed stock; 0.55 = 6/11 for

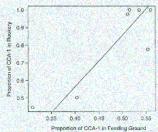


Fig. 2. Relationship between the frequency of haplotype CC-A1 in seven juvenile freeding areas and seven adjacent nesting boaches. The resulting correlation ($R^2 = 0.88$) was significant at P = 0.049.

Northern, 0.09 = 1/11 for Southern, and 0.27 = 3/11 for Gulf (the denominator of 11 includes 10 feeding populations plus the 'other' category, not shown). Furthermore, the 2.5 and 97.5 percentiles of the posterior contribution for many of the stocks ranged from nearly zero to a proportional contribution above 0.5. In fact, none of the rookeries alone showed 'significantly' disproportionate contributions to the closest lumped mixed stocks, where we define significance as a 2.5 percentile of the relative contribution greater. than 0.53, 0.09, or 0.27, respectively (according to the partitioning discussed above). Nevertheless, there were patterns in the results: the NEFL-NC appeared to contribute disproportionately to the Northern mixed stock (median 0.77, 2.5 percentile 0.498, null expectation 0.55), and South Florida contributed slightly more to the Gulf mixed stock (median 0.33, 2.5 percentile 0.13, null 0.25). In the disaggregated results (treating each feeding area separately; data not shown), structure was reflected in slightly enhanced contributions (> 10% over an expected proportional contribution of 9.1%) from South Florida to TX, FL-SG, GA and VA: from Mexico to FL-NG, FL-SA and SC; and from NEFL-NC to NC, VA and NE-US (see Tables I and 2 for abbreviations).

Looking at the results in a more traditional way, as the fraction of each feeding ground population contributed by specific rookeries (Table 5), the estimated contributions were (as expected) dominated by the size of the contributing rookeries: the three aggregate feeding ground populations were estimated to have 82–90% contributions from the large rookery in south Florida. The man conclusion to draw from 161be 5 is the very small contribution of mokeries

Table 4 Results of mixed stock analysis: proportions from each rookery contributed to specified feeding grounds

Raokery	Feeding ground	Mean	SD	2.50 percentile	Median	97.50 percentili
FL-NG	North	0.6202	0.1449	0,3209	0,6302	0.8707
	South	0.08726	0.07969	0,002518	0,06432	0.2973
	Gulf	0.2926	0.1376	0,07244	0,2769	0.5909
SFL	North	0.6058	0.1203	0.3674	0.6705	0.8255
	South	0.0555	0.03944	0.01028	0.04359	0.1596
	Gulf	0.3387	0.118	0.129	0.3322	0.578
MX	North	0.4828	0.1229	0.2498	0.4827	0.7207
	South	0.2091	0.1019	0.03463	0, 1943	0.4441
	Gulf	0.3081	0.1148	0.1127	0.3001	0.5535
NEFLINC	North	0.7514	0.1114	0.498	0.7653	0.9262
	South	0.02831	0.0299	7.40 × 10 ⁻⁴	0.01916	0.3077
	Gulf	0.2203	0.1095	0.05372	0.205	0.4723
DT	North	0.5988	0.1463	0.3026	0.6049	0.8596
	South	0.1092	0.09249	0.002652	0.07686	0.3451
	Gulf	0.298	0.1368	0.07421	0.284	0.5925
GR	North	0.5985	0.1477	0.2962	0.6051	0.8621
	South	0.09958	0.08974	0.002792	0.07463	0.3436
	Gulf	0.3019	0.1386	0.07637	0.2885	0.603
HR	North	0.6071	0.1464	0.3076	0.6147	0.8666
	South	0.08433	0.08125	0.002017	0.05967	0.3047
	Gulf	0.2083	0.1399	0.07803	0.2952	0.6105
TR	North	0.5934	0.1484	0.2947	0.5998	0.8611
	South	0.1009	0.09136	0.002971	0.07512	0.3409
	Gulf	0.3057	0.139	0.00793	0.2921	0.6086

Abbreviations: FL NG, northwest Florida; SFL, south Florida; NEFL NC, northeast Florida to North Carolina; DF, Ory Toriugas, MX, Mexico; BR, Brazzl; GR, Greece; TR, Turkey.

outside the northwest Atlantic (Greece, Brazil, and Turkey). Despite their non-negligible size, we showed that these rookeries provided at most (95% confidence limit) about 1% of the turties in our foral mixed stucks. Since Table 4 shows the partitioning among mixed stocks of the total contribution to northwest Atlantic feeding habitats, it cannot provide this information).

The nesting colonies designated here as source populations represent most (but not all) of the known nesting effort in the Atlantic-Mediterranean system (Ehrhart et al. 2003; Margaritoulis et al. 2003). Important nesting efforts occur in Cuba. Cape Verde Islands, and along the coast of Africa, but could not be included for a variety of logistical reasons. Furthermore, additional nesting colonies may await discovery in undersurveyed regions. It is important to remember these limitations when formulating an interpretation of mixed stock analyses. However, more than 99% of the haplotypes observed in juvenile populations (excepting CC-A18, CC-A19, CC-A22, CC-A23; n = 4) could be matched to haplotypes in nesting populations. providing at least a qualitative assurance that most of the genetic diversity was captured in the existing rookery samples.

Discussion

Previous investigations have revealed that loggerhead turtles may cross entire ocean basins during their posthatchling phase (Bowen et al. 1995; Bolten et al. 1998; Resendiz et al. 1998; Nichols et al. 2000; Alfaro-Shigueto et al. 2004). Juveniles from nesting beaches in the northwest Atlantic inhabit oceanic zones around the Azores, Madeira, Grand Banks (Newfoundland, Canada), off the coast of Africa, and throughout the Mediterranean Sea (Bolten 2003a). Laurent et al. (1998) demonstrated that about half of the oceanic-stage juveniles in the Mediterranean originated on beaches of the western Atlantic. At the same time, the older neritic-stage turtles in the Mediterranean included little or no contribution from the western Atlantic (Laurent et al. 1998). In other words, by the time these Atlantic loggerheads switch from pelagic to peritic feeding, they have departed the Mediterraneon and reappear in continental shelf habitats on the other side of the Atlantic, Notably, the switch from oceanic to peritic stages is not immutable, as both older juveniles and adults can return to oceanic habitats (Eckert. & Martins 1989: Hatase et al. 2002b: Witzell 2002: Bolten

Table 5 Results of mixed stock analysis: proportions of each feeding ground contributed by specified rookeries

Feeding ground	Rookery	Mean	SD	2.50 percentile	Median	97.50 percentile
North	FLING	0.008292	0.002842	0.003652	0.00797	0.0148
	SFL.	0.8645	0.03129	0.7909	0.8687	0.9139
	MX	0.01947	0.006148	0.009428	0.01882	0.08353
	NEFL-NC	0.1018	0.02579	0.0592	0.0989	0.1614
	DT	0.002898	0.00101	0.001259	0.00279	0.005179
	GR	0.001355	0.00155	7.91 × 10-5	0.001094	0.005711
	BR	9.80 × 10→	8.82×10-	5.57 × 10 · 5	7.34 × 10-4	0.003294
	TR	5.70 × 10 4	5.54 × 10→	3.11 × 10 ⁻⁵	4.11×10 4	0.002044
South	FL-NG	0.01674	0.01959	0.61 × 101	0.01038	0.07155
	SFL	0.8215	0.07938	0.6262	0.8352	0.9367
	MX	0.103	0.04618	0.03393	0.09557	0.2122
	NEFL-NC	0.0442	0.03909	0.001333	0.03336	0.1458
	DT	0.007729	0.01019	1.31 × 10 1	0.004337	0.03522
	GR	0.003635	0.00636	2.71 × 10-5	0.001512	0.02036
	BR	0.001733	0.002521	1.51 × 10-1	8.27 × 10-4	0.008925
	TR	0.001478	0.002734	9.97 × 10-4	5.64 × 10-4	0.008591
Gulf	FL-NG	0:006005	0.005371	0.001609	0.00677	0.02194
	SFL	0.9022	0.04311	0.7959	0.9104	0.9611
	MX	0.02523	0.0133	0.007518	0.02252	0.05865
	NEFL NC	0.05827	0.03239	0.01363	0.05253	0.1372
	DT	0.002963	0.001972	5.90 × 10-+	0.002512	0.007999
	GR	0.00165	0.002143	5.66×10-4	9.87 × 10 →	0.007048
	BR	0.001119	0.001428	3.48 × 10 ÷	6.51 × 10 →	0.005034
	TR	6.29 × 10→	8.21 × 10-4	2.08 × 10-9	3.76 × 10 -4	0.002792

NEFL NC is a category combining neating beaches from northeast Florida, Georgia, South Carolina and North Carolina. Other abbreviators are described in Tables 1-4.

Three analyses were conducted to test for population subdivisions and natal homing behaviour in juvenile loggerhead turties of the northwest Atlantic: (i) an AMOVA for juvenile feeding populations ($\Phi_{e\tau} = 0.0088$; P = 0.016); (ii) correlation statistics to compare genotype frequency differences in nesting populations vs. adjacent feeding populations (Mantel $R^2 = 0.52$; P = 0.001), and (iii) a mixed stock analysis using Bayesian methodology. All three approaches indicate a nonrandom distribution of juvenile turtles, and a significant relationship between nesting colonies and adjacent feeding populations. Collectively these analyses yield substantial evidence of natal homing in developmental migrations. Tagging studies, indicating high site fidelity in juvenile turtles on the Atlantic coast, indirectly support this conclusion (Avens et al. 2003; Hopkins-Murphy et al. 2003).

One cavest to these conclusions is that the correlation statistics are based on a subset of the entire database, agecifically the seven nesting locations along the continental coastline of the southeastern USA (Table 1), and their adjacent feeding populations (Table 1). A second caves it is that sampled individuals may have included a few small adults, and certainly included a wide range of juvenile age classes, from new nertitic-stage recruits to older turtles approaching maturity. A third qualification is that we have not sampled the entire range of feeding habitait in the northwest Atlantic (see Engstrom et al. 2002). Given these limitations, we caution that the affinity described here between nesting colonies and adjacent juvenile feeding areas does not verifiably extend beyond the continental shelf of North America.

Population structure and life history implications

Subsequient to the coemic juverille singe, loggeriead turtles switch to primarily neritic, benthic foraging habitats. These neritic foraging habitats may be a great distance from the occunic habitats, from floja California to Japan, for example, or from the Azores to the eastern coast of North America. Whereas the migration to oceanic feeding areas is episcently facilitated by passive drift, the return trip may include active swimming. Once juveniles return to their region of origin and switch to benthic feeding, they may occasionally return to a pelagic feeding mode, as indicated by satellite telemetry, stable isotope ratios and tag returns (Eckort & Martine 1969, Hatase et al. 2002b), Juvenile truther make & Martine 1969, Hatase et al. 2002b).

seasonal migrations into temperate habitats (such as the northeast US coast), and adults make reproductive migrations over hundreds of kilometres (Limpus et al. 1992; Schroeder et al. 2003). We conclude that the complex like history of loggerhead turles may include two homing migrations. The first is a unigration from oceanic habitat (often thousands of kilometres from the nesting beach) to the region of origin. The second is the cyclic reproductive migration from adult foraging habitat to courting grounds and nesting habitat.

Our analyses demonstrate that there is genetic structure among feeding ground populations and that this genetic structure is spatially correlated with the genetic structure of nesting populations. The technique that comes closest to answering our specific question-how the contribution of rookeries to foraging grounds is partitioned-results in wide confidence intervals, but with some consistent trends. First, there is little contribution from assayed nesting colonies outside the northwest Atlantic (the parameter giving relative contributions from outside the northwest Atlantic has a mean of 0.03, with an upper confidence limit of 0.11). Second, there are indications (supporting our simpler analyses of correlation in frequency of the dominant haplotype) of targeted contributions from rookeries to nearby feeding grounds, especially from the south Florida rookery to foraging grounds in the Gulf of Mexico and from Atlantic rookeries (NEFL-NC) to Atlantic foraging habitats. However, no rookery-foraging ground pair actually shows significantly greater contributions than a proportional null model. Mixed stock analyses do not allow strong conclusions at this level of resolution. While this limitation can partly be overcome with more data, some of it is inherent in the overlap of haplotype profiles among rookeries and foraging grounds. We are working to incorporate spatial structure into the framework of mixed stock analysis, so that we can more powerfully test specific spatial hypotheses (Bolker et al. unpublished results).

Marine furthes have a complex population structure, with lower levels of population differentiation in nucleus with lower levels of population differentiation in nucleus DNA assays (Karl et al. 1992; FirstSimmons et al. 1996; 1997b; Peance 2001; Roberts et al. 2004; Superminosed on this pattern are the life history stages of loggerhead turtles, with varying degrees of population structure. Oceanic juveniles are well mixed in the North Atlantic (Rotten et al. 1998; LaCassella et al. 2004; P. Dutton unpublished data); the nortic juveniles of the west-mathantic subsequently segregate at all low but significant level ($\Theta_{\rm eff} = 0.0088; P = 0.016);$ and corresponding neating populations are highly structured ($\Theta_{\rm hf} = 0.27; P < 0.001$ for the seven nowheries compared to adjacent feeding cohocts).

In this study we consider two primary influences on the distribution of nerific-stage juveniles; the size of the source populations and the proximity of juvenile feeding habitat to these source populations. However, additional factors will certainly influence the distribution of juveniles. Hopkins-Murphy et al. (2003) demonstrate that larger juveniles dominate the feeding habital in the eastern Golf of Mexico GE-NG and FL-SG), while smaller juveniles are more prevalent in the pertpheral and seasonal habitats of the western Gulf of Mexico (CN) and the northerast USA. Analyses or hapfolype distributions among size classes may prove fruitful in teasing apart these additional life-history components.

Notably, one life stage in North Atlantic loggerheads remains to be evaluated with intDNA surveys: the adult feeding populations. Little is known about sduft feeding palaties, but (for loggerheads nesting in the southeast DNA) new principles size along the east coast of the USA, the Bhaanas, Cuba, Culf of Mexico and Caribbean Mexico Chopkins Murphy et al. 2003; Schroeder et al. 2003. It will be informative to survey the adult enhors and determine whether they also segregate on freeling grounds, it would also be informative to test the pivernile knowing hypothesis in other regions (Mediterranean Ses, Japan), and in other son turtle speeds.

Conservation implications

The finding of significant population structure in juvenile loggerhead turtles carries some implications for wildlife management. The hazards that affect breeding populations may also have an impact on the next generation that is feeding in nearby waters. However, homing is not absolute and considerable movement occurs. as well. The stranding data from Texas and the northeast USA, where nesting is sparse or absent, illustrate that feeding populations extend far past the regional nesting habitat (Fig. I). One consequence of this widespread foraging is that juvenile turtles originating in Yucatan Mexico are feeding in US waters (Table 5). We suspect the converse is true (Table 4). This finding invokes the provisions of the 1982 United Nations Convention on the Law of the Sea, in which nations that host the developmental habitat for migratory marine species hold fishing rights for these animals on the high seas (Van Dyke 1993). The 1983 UN Convention on the Conservation of Migratory Species (also know as the Bonn Convention) prohibits taking endangered species during migrations on the high scas (Hykle 1992). Under the principles outlined in these international agreements, nations that host nesting and developmental habitats for marine turtles have some level of jurisdiction over these animals on geographically remote feeding grounds, even if those feeding grounds are within the territorial boundaries of another nation. Activities in US waters can deplete an embattled rookery in Mexico, and activities in Mexico could have an impact on threatened nesting populations in the southeastern USA. Provisions of international law apply here.

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